

## Protocol and budget for real-time PCR diagnostics for honey bees

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Sketched out below are the technical details for a routine we use regularly to generate genetic data from honey bee samples. These samples range from field collections of adult or larval bees to controlled experiments carried out on adult bees. The protocol focuses on the RNA of collected samples, a route that is essential for detecting the many bee viruses and one that also provides an opportunity to assess activity levels of specific honey bee or pathogen genes.

### Acid Phenol RNA Extraction

#### *Needed Reagents:*

- Lysis buffer: 0.8M Guanidine thiocyanate, 0.4M Ammonium thiocyanate, 0.1M Sodium acetate, 5% glycerol, 2% Triton-X100. For 620 ml: 94.53g Guanidine thiocyanate, 30.45g Ammonium thiocyanate, 33.4 ml 3M stock Sodium acetate, 50 ml glycerol, 20 ml Triton-X100, Read MSDS on thiocyanate safety. Mix in 300 ml water, bring up to 620 ml then autoclave. Use within 30 days.
  - Acid Phenol (pH 4, Sigma Aldrich)
  - Chloroform
  - Isopropyl alcohol
  - 70% ethanol
  - Sterile, DNase/RNase free water
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- Collect ca. 200 bees from a field colony, or 30-50 from a cage experiment (Fig. 1A), into a 50 disposable centrifuge tube and flash freeze in liquid nitrogen, on dry ice, or in a -80°C freezer
  - Count 50 bees from tube and place in a sealable bag (1 quart Zip-Loc or BioReba, [www.bioreba.ch](http://www.bioreba.ch), sample extraction bag; Fig. 1B)
  - Add 25 ml (or 500 µl/bee) Lysis buffer
  - Seal bag w/o air and smash with rolling pin until bees broken fully, for 30 sec (Fig. 1C)
  - Massage liquid to mix fully, then draw off 620 µl into a labeled microcentrifuge tube, avoiding bubbles
  - Freeze this aliquot immediately at -80°C
  - Thaw sample briefly, while still slightly frozen add 380 µl acid phenol, vortex 30 s, incubate at 95°C for ten min

- Cool to RT (can do on ice 5-10 min) and add 200  $\mu$ l chloroform
- Shake tubes vigorously by hand for 15 sec and then incubate at room temperature (RT) for 2 to 3 min.
- Centrifuge the samples at  $10,000 \times g$  for 15 min at  $4^{\circ}\text{C}$ .
- Collect upper aqueous phase into fresh tube containing 500  $\mu$ l isopropyl alcohol, vortex 30 sec. (Can stop at this point, sample is stable for days at  $4^{\circ}\text{C}$ )
- Incubate samples at  $4^{\circ}\text{C}$  for 30 min and centrifuge at  $10,000 \times g$  for 10 min at  $4^{\circ}\text{C}$ .
- Remove the supernatant. Wash the RNA pellet once with 1 ml 70% ethanol
- Mix the sample by vortexing and centrifuge at  $7,000 \times g$  for 5 min at 2 to  $8^{\circ}\text{C}$ .
- Pour off supernatant, centrifuge 30 sec and pull off remaining supernatant with pipette
- Briefly dry the RNA pellet (air-dry 5-10 min).
- Dissolve RNA in 50  $\mu$ l RNase-free water by incubating for 10 min at 55 to  $60^{\circ}\text{C}$ .
- Store at  $-80^{\circ}\text{C}$

### **Complementary DNA (cDNA) synthesis**

#### *Needed Reagents:*

DNase I (2 U/ $\mu$ l solution)

RNaseOut (Invitrogen)

dNTP (100mM solution)

DNase I Buffer (10X)

Random primer set (7-mer at 10 mM concentration)

Superscript II enzyme (200 U/ $\mu$ l; Invitrogen)

Superscript Buffer (10X)

DTT (0.1 M)

- Prepare initial reaction mix containing, for each sample, 1  $\mu$ l DNase I solution, 0.5  $\mu$ l primer mix, 0.4  $\mu$ l RNaseOut, 0.4  $\mu$ l dNTP mix, 1.1  $\mu$ l DNase buffer.
- Aliquot 2.9  $\mu$ l reaction mix per well
- Add 8  $\mu$ l of 1  $\mu\text{g}/\mu\text{l}$  total RNA per well
- Incubate  $37^{\circ}\text{C}$  1 hour, then  $75^{\circ}\text{C}$  for 10 min, cool to  $4^{\circ}\text{C}$
- Heat plate to  $70^{\circ}\text{C}$  10 min, place on ice
- When cooled, centrifuge briefly
- Incubate  $42^{\circ}\text{C}$  for 2 min
- Add 4  $\mu$ l Superscript mix to each sample (0.5  $\mu$ l Superscript II enzyme, 1.5  $\mu$ l Superscript buffer, 2  $\mu$ l DTT)
- Incubate  $42^{\circ}\text{C}$  50 min,  $70^{\circ}\text{C}$  15 min,  $4^{\circ}\text{C}$

## Quantitative PCR (20 µl reaction)

### *Needed Reagents:*

Sterile H<sub>2</sub>O

Specific Forward and Reverse oligo primers

Express SYBR Green ER qPCR SuperMix Universal (Buffer, Taq polymerase, dNTP's)\*.

\*It is of course common to mix qPCR reagents by hand, rather than buying a prepared mix, and numerous published papers describe such reaction mixes (e.g., one 'home-made' SYBR Green recipe that we have used for several years is described in Evans, 2006).

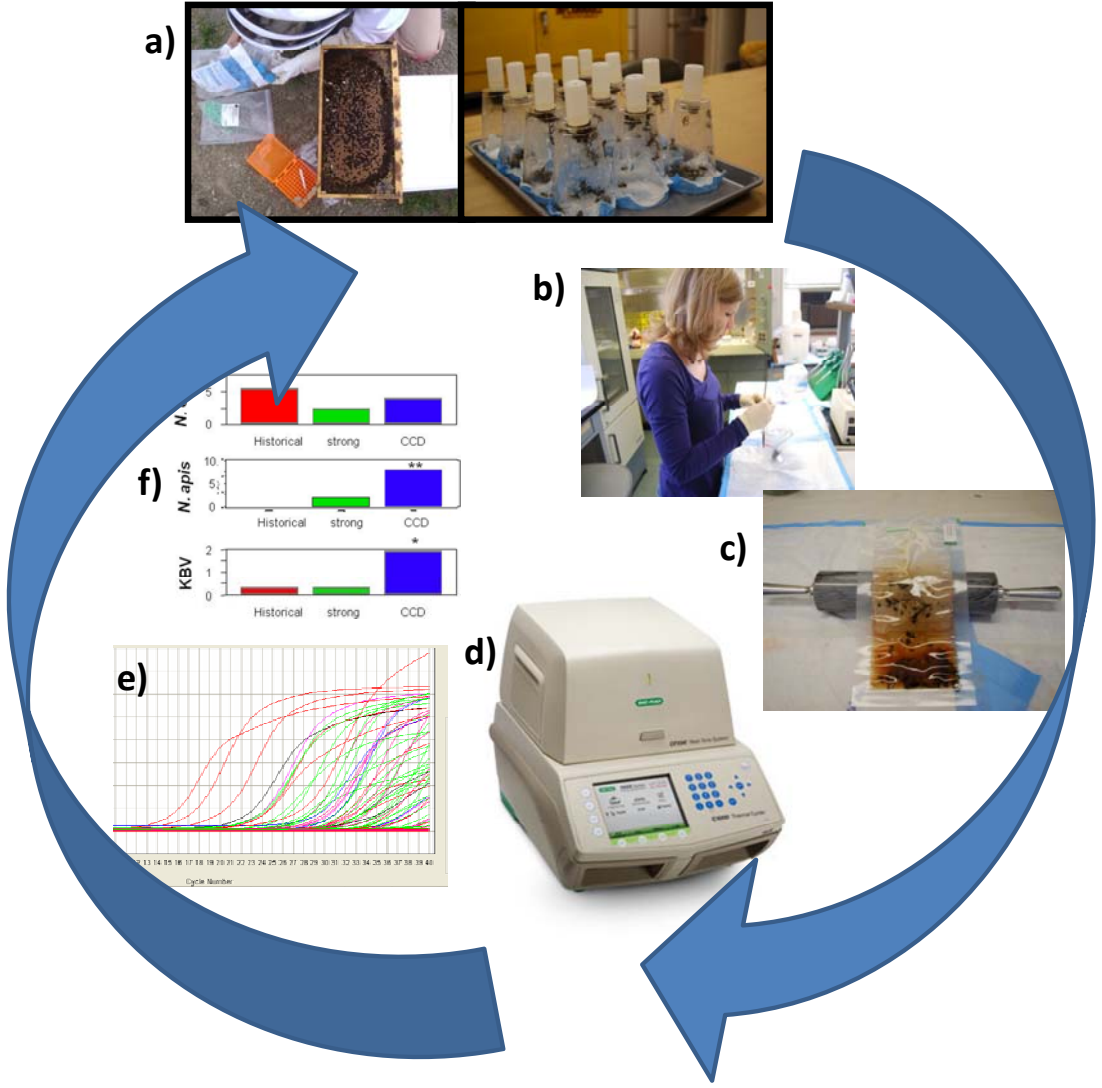
- Prepare Master Mix for each sample plus handling excess (calculate based on 100 samples for 96-well plate)
- Aliquot into wells of plate, 19 µl each on ice
- Add 1 µl template per well, on ice
- Centrifuge plate to incorporate sample and remove bubbles
- Program thermal cycler\*\*, add plate, and run program (Fig. 1D)
- Collect post-run data for spreadsheet manipulation and statistics (Fig. 1E, 1F).

\*\* We use a Bio-Rad CFX-96 thermocycler, programmed to start with a 50°C two minute step (this safety step enables the degradation of any possible past PCR products since the SuperMix incorporates dUTP into all PCR products and also includes Uracil DNA Glycosylase which will degrade any pre-existing uracil residues). Next is a 2 minute 95°C denaturing step, followed by 40 cycles of 95°C for 20 seconds, 60°C for 30 seconds, 72°C for 80 seconds. The fluorescence is measured at the 72°C step, since this provides another barrier against measuring short non-specific primer products (Evans, 2006).

## Reference

Evans JD (2006) Beepath: An ordered quantitative-PCR array for exploring honey bee immunity and disease.

**Figure 1. Schematic showing key steps in isolation of RNA and diagnostics using quantitative PCR (see text).**



**Rough Budget Estimate for quantitative PCR on seven bee targets plus one control gene  
(U.S. Dollars, Jan. 2011)**

<b>Sample Prep</b>	Price/Unit	per Unit	Units/sample	Price/Sample
RNA extraction	\$200/96	2.08	1	2.08
Superscript II	\$800/10000	0.08	50.00	4.00
Random 7mers	.0025/rxn	0.0025	1.00	0.0025
Disposable Plates	\$220/200	1.1	0.01	0.01
Total Extraction/cDNA				<b>6.10</b>

(per sample prep)

**Quantitative PCR**

	Price/units	per Unit	Units/Reaction	Price/Reaction	Price/8 Targets
SuperMix Reagent	1506/2000	0.75	1.00	0.753	6.02
Primer1	\$5/2000	0.0025	1.00	0.0025	0.02
Primer2	\$5/2000	0.0025	1.00	0.0025	0.02
PCR Plate/cover	\$650/100	6.50	0.01	0.065	0.52
Total				0.83	<b>6.58</b>

**Labor:** ca. 20 samples/person-day (trained undergraduate assistant)

\$16.33/hr x 8 =	130.64/20	<b>6.53</b>
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(per sample Labor)

<b>per sample labor and supplies combined:</b>	<b>19.23</b>
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<b>Project with 600 samples x 8 targets</b>				
	<b>RNAprep</b>	<b>PCR</b>	<b>Labor</b>	<b>TOTAL</b>
<b>Total Cost</b>	<b>3658.38</b>	<b>3948</b>	<b>3919.20</b>	<b>11525.58</b>